

Acclimation training improves endurance cycling performance in the heat without inducing endotoxemia

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Submitted to Journal:
Frontiers in Physiology

Specialty Section:
Exercise Physiology

ISSN:
1664-042X

Article type:
Original Research Article

Received on:
25 Apr 2016

Accepted on:
13 Jul 2016

Provisional PDF published on:
13 Jul 2016

Frontiers website link:
www.frontiersin.org

Citation:

Guy J, Pyne DB, Deakin G, Miller CM and Edwards AM(2016) Acclimation training improves endurance cycling performance in the heat without inducing endotoxemia. *Front. Physiol.* 7:318. doi:10.3389/fphys.2016.00318

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Provisional

1 **Title: Acclimation training improves endurance cycling**
2 **performance in the heat without inducing endotoxemia.**

3
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21 **Abstract**

22 **Purpose:** While the intention of endurance athletes undertaking short term heat training
23 protocols is to rapidly gain meaningful physical adaptation prior to competition in the heat, it is
24 currently unclear whether or not this process also presents an overt, acute challenge to the
25 immune system. The aim of this study was therefore to examine the effects of heat training
26 on both endurance performance and biomarkers associated with inflammatory and immune
27 system responses. **Methods:** Moderately-actively males (n=24) were allocated randomly to
28 either HOT (n=8, 35°C and 70% RH; NEUTRAL (n=8, 20°C and 45% RH); or a non-
29 exercising control group, (CON, n=8). Over the 18 day study HOT and NEUTRAL
30 performed seven training sessions (40 min cycling at 55% of $\dot{V}O_2$ max) and all participants
31 completed three heat stress tests (HST) at 35°C and 70% RH. The HST protocol comprised
32 three x sub-maximal intervals followed by a 5 km time trial on a cycle ergometer. Serum
33 samples were collected before and after each HST and analysed for interleukin-6,
34 immunoglobulin M and lipopolysaccharide. **Results:** Both HOT and NEUTRAL groups
35 experienced substantial improvement to 5 km time trial performance (HOT -33 ± 20 s, $p =$
36 0.02 , NEUTRAL -39 ± 18 s, $p = 0.01$) but only HOT were faster (-45 ± 25 s and -12 ± 7 s, p
37 $= 0.01$) in HST₃ compared to baseline and HST₂. Interleukin-6 was elevated after exercise for
38 all groups however there were no significant changes for immunoglobulin M or
39 lipopolysaccharide. **Conclusions:** Short-term heat training enhances 5 km cycling time trial
40 performance in moderately-fit subjects by ~6%, similar in magnitude to exercise training in
41 neutral conditions. Three top-up training sessions yielded a further 3% improvement in
42 performance for the HOT group. Furthermore, the heat training did not pose a substantial
43 challenge to the immune system.

44

45 **Key words:** cycling, heat acclimation, inflammation, lipopolysaccharide, cytokine, endurance
46 performance

47 **Introduction**

48 Short- and medium-term heat acclimation training protocols are widely used by endurance
49 athletes to increase both heat tolerance and subsequent competitive performances in the heat
50 (Périard, Racinais, and Sawka. 2015). Although favourable performance and physiological
51 benefits can be realized from short term programs (≤ 7 days), greater benefits are likely from
52 longer protocols (7-14 days) (Daanen et al, 2011; Guy, et al. 2015; Lorenzo et al. 2010;
53 Nielsen et al. 1997). For elite athletes, busy training and performance schedules limit the time
54 is available for strategies such as heat training, and addition of supplementary training
55 sessions may sustain and/or complement the initial adaptations.

56 While the acute effects of short-term heat exposure on blood biomarkers associated with
57 inflammation have been reported (Gill et al. 2015; Hailes et al. 2011), few studies have
58 investigated the effects of longer duration heat training. The human immune system can
59 usually deal with mild-to-moderate inflammatory responses, however, when a heat stimulus
60 is too large, systemic inflammation can result in heat shock and potentially fatal sepsis
61 (Bouchama et al. 2007). Athletes will generally seek a heat training stimulus that is large
62 enough to evoke a training adaptation; however, there likely comes a point where the risk of
63 clinical or subclinical levels of immune disturbance increases.

64 Exercise-induced endotoxemia is a potential risk of strenuous activity in the heat primarily
65 attributed to translocation of lipopolysaccharide (LPS) from the gut into the circulation (Lim
66 et al. 2009). An abundance of circulating LPS can evoke an inflammatory response, leading
67 to heat shock and overwhelming anti-LPS mechanisms including immunoglobulin M (IgM)
68 (Camus, et al. 1998) and cytokines operating in an anti-inflammatory role such as interleukin-
69 6 (IL-6) (Abbasi et al. 2013). Consequently, when anti-LPS mechanisms and rate of LPS
70 clearance are inadequate to counter the heat-induced increase of LPS, endotoxemia may
71 ensue. This outcome could potentially occur during a period of heat acclimation training if
72 the athlete is unable to cope with the thermal loads presented. As IgM is a key antibody in
73 neutralising LPS (Camus et al., 1998), its concentration in circulating blood can reflect the
74 body's response to endotoxin accumulation, and the degree of protective capacity in the event
75 of further challenges. IgM concentration can increase substantially (~20%) after exercise in
76 the heat, although this elevation does not occur following five days of heat training (Hailes, et
77 al. 2011). Of the few studies that have investigated IL-6 as a blood biomarker during
78 exhaustive exercise in the heat, Selkirk and colleagues (2008) observed a twenty-fold
79 increase in plasma concentrations following 2 h of exhaustive walking in protective clothing
80 in very hot and humid conditions, with IL-6 inhibiting endotoxin induced increases in tumor
81 necrosis factor alpha and cytokines. Furthermore, the neuroinflammatory response to exercise
82 indicates that an increase in cytokine concentration such as IL-6 reaching a critical threshold,
83 it is likely that sensations of fatigue develop to prevent traumatic injury of specific organs
84 and other physiological systems within the body (Vargus & Marino, 2014) Therefore,
85 athletes who undertake short or medium duration heat acclimation training programs could
86 potentially be at increased risk of exercise-induced heat stress and immune disturbances
87 associated with fatigue.

88 Recreationally-active healthy adults often participate in one-off events such as an ironman
89 triathlon, marathon and week-long sporting events such as the Masters' Games. It appears
90 that the threshold for the onset of exercise-induced endotoxemia is lower in untrained than
91 trained individuals (Selkirk et al. 2008). Individuals seeking to use heat acclimation training
92 as an additional training stimulus may choose either a short- or medium-term program, to
93 elicit the classic thermal markers of plasma volume expansion, lower heart rate at

94 submaximal intensities and lower end point core temperature, which collectively promote
95 aerobic performance (Guy et al, 2015). However addition of a heat load to training can often
96 be very demanding, with some studies implementing challenging protocols on their
97 participants, e.g. 90 min of cycling for 10 consecutive days (Gibson et al. 2015). It is prudent
98 to account for both training load and accumulated inflammation from heat stress over the
99 training period. As longer heat training sessions (>60 min) are likely fatiguing for
100 recreationally-trained athletes, and can increase peripheral fatigue compared with shorter
101 protocols (Wingfield et, 2016), the addition of shorter and supplementary training sessions
102 could yield similar benefits, but with lower overall stress.

103 Few studies have investigated the degree of inflammation and endotoxemia associated with
104 short- and medium-term heat acclimation training. Therefore, the aim of this study was to
105 investigate whether short-term heat training with the addition of supplementary sessions can
106 improve cycling time trial performance (TT), improve sub-maximal exercising heart rate and
107 core temperature, and to quantify the degree of inflammation associated with heat
108 acclimation training.

109

110 **Methods**

111

112 *Design*

113 This study consisted of three groups of recreationally-active male athletes: a heat training
114 group (HOT), a matched thermo-neutral training group (NEUTRAL) and a control (no
115 training) group (CON), in a pre-post parallel groups design.

116

117 *Participants*

118 Twenty four moderately trained male participants (3 ± 1 moderate-high intensity training
119 sessions per week, duration 60 ± 15 mins; mean \pm SD) aged 24.5 ± 3.8 years, height 178 ± 7
120 cm, mass 84.6 ± 10.8 kg, body fat $17.5 \pm 6.1\%$, and maximal oxygen uptake ($\dot{V}O_2$ max) of
121 45.0 ± 5.0 ml.kg.min⁻¹ volunteered for the study. Prior to taking part, participants provided
122 written informed consent in accordance with the Declaration of Helsinki and underwent a
123 pre-screening health questionnaire including use of anti-inflammatory or immunomodulating
124 medications (none were present). The study protocol was approved by the James Cook
125 University Human Research Ethics Council (Approval number H5647).

126

127 *Methodology*

128 Assessment of $\dot{V}O_2$ max was undertaken on a cycle ergometer (VeloTron and Velotron
129 Coaching Software, Racermate, United States) at least 72 h before beginning the
130 experimental trials. The intervention comprised a short-term training protocol of four training
131 sessions on consecutive days, followed by three supplementary training sessions every three
132 days. All participants completed three heat stress tests (HST₁₋₃) and seven training sessions
133 over 18 days, with HST₁ performed as a baseline measure of heat tolerance, HST₂ completed
134 between the end of the short-term program and before beginning the supplementary top-up
135 training, and HST₃ completed 48 hours after the final supplementary training session (Figure
136 1). Each group performed the HST in a custom-built environmental chamber at a temperature
137 of 35°C and 70% RH. Participants in the HOT and NEUTRAL conditions completed exercise
138 training sessions in hot and humid (35°C and 70% RH) or thermo-neutral conditions (20°C
139 and 50% RH) respectively. Participants in the CON group did not undertake exercise training
140 but completed the three HST's at the same intervals as HOT and NEUTRAL groups.

141 Participants were instructed to rest and avoid moderate or high levels of physical activity on
142 days that they were not required to attend the laboratory.

143

144 *Test of Maximal Oxygen Uptake*

145 Maximal oxygen uptake was determined by an incremental test to exhaustion on a cycle
146 ergometer (VeloTron and Velotron Coaching Software, Racermate, United States). Briefly,
147 the test began with participants cycling at 80-90 rpm at 120 W, with the workload increasing
148 by 20 W every min until volitional exhaustion or when cadence was unable to be maintained
149 above 80 rpm. Expired gases were collected via a one-way breathing system (Hans-Rudolph,
150 United States) and analysed by a calibrated Moxus Metabolics Measurement cart (AEI
151 Technologies, United States). Attainment of $\dot{V}O_2$ max was determined by the satisfaction of
152 standard criteria (Midgley et al. 2007).

153 *Heat Stress Test*

154 The heat stress test was of similar design to earlier work (Garrett et al. 2009; Lorenzo et al.
155 2010) and comprised cycling for three x 10 min submaximal workloads with a 3 min rest
156 period between workloads, followed by a 5-km self-paced time trial (TT). Following a 5 min
157 standardised warm-up, the participants completed three 10 min workloads at 50%, 60% and
158 70% of their peak wattage corresponding to their individualised $\dot{V}O_2$ max. After the 70%
159 workload was complete, a 5 min rest period was given before the start of the TT. Participants
160 were able to view their rpm and were informed of the distance travelled every 500 m to assist
161 with pacing. Heart rate (RS400, Polar Elektro, Finland), and core temperature (T_c) (ttec 501-3
162 data logger and data logger software version 10.1, Nordex Pty Ltd, Australia; MEAS 449 1RJ
163 rectal temperature thermistor, Measurement Specialities, United States) were sampled at 5 s
164 intervals. Fluid intake (water, *ad libitum*), rating of perceived exertion (Borg RPE 6 – 20,
165 Borg 1970) and thermal comfort (TComf) were recorded throughout the test. Nude dry body
166 mass was recorded pre and post exercise on a calibrated set of scales (BF-522W, Tanita,
167 Japan) and body mass was adjusted for fluid loss and expressed as a percentage change.

168

169 *Blood collection*

170 Upon arrival at the laboratory, participants rested for 20 min before blood collection was
171 performed. Blood was drawn in a seated position 10 min before and 10 min after each HST
172 via a 22g needle from a prominent superficial forearm vein located at the antecubital fossa,
173 and drained directly into an 8.5 ml sterile serum separator Vacutainer tube containing a clot
174 activator and gel for serum separation (Beckton and Dickson, USA). Samples were
175 refrigerated at 4°C for 30 min to allow clotting and then centrifuged at 1000 x g at 6°C for 10
176 min (Rotina 420R, Hettich, Germany). Serum was removed and stored in 400 μ l aliquots that
177 were frozen immediately for a maximum of three months at -80°C for later analysis. Serum
178 concentrations of IL-6 (Quantikine HS600B, R&D Systems, United States), IgM (AB137982,
179 Abcam PLC, United Kingdom), and LPS (HIT302, Hycult, Biotechnology, Netherlands)
180 were analysed in duplicate by ELISA according to manufacturer's instructions.

181 *Aerobic Interval Training*

182 Participants in HOT and NEUTRAL undertook matched aerobic interval training on a cycle
183 ergometer (Monark Ergomedic 828 E, Sweden) in hot and humid (35°C and 70% RH) or
184 thermo-neutral conditions (20°C and 50% RH) respectively. The exercise-training
185 intervention included seven training sessions comprised a standardised 3 min warm-up
186 followed by 4 x 10 min interval at a fixed workload of 55% $\dot{V}O_2$ max. A three min rest period
187 was given between each workload and water consumed *ad libitum*. A shorter duration
188 interval-based protocol was chosen to better reflect the training status of the recreationally-
189 trained participants; interval-based training has been shown to be beneficial for heat
190 acclimation (Dawson et al. 1989; Kelly et al. 2016), and shorter duration training can reduce
191 cumulative fatigue (Wingfield et al. 2015) while promoting performance (Nielsen et al 1997).

192 Heart rate was recorded at 5 sec intervals and RPE and TComf recorded at the end of each
193 interval. Participants self-reported symptoms of illness, inflection, soreness or inflammation
194 prior to the start of each training session. No symptoms of illness or infection were reported.

195 ***Figure 1 about here***

196

197 *Statistical Analysis*

198 Data that passed tests for homogeneity of variance were analysed by a mixed-model analysis
199 of variance or t-test (where appropriate) and significance accepted when $p \leq 0.05$. Where
200 significant differences were indicated they were identified with the *post hoc* Tukey Test. Data
201 is expressed as mean \pm SD and change scores expressed as mean \pm 90% confidence limits
202 (CL). The baseline TT performance (s) was not normally distributed and therefore analysis of
203 covariance was used to investigate between-group differences with participant $\dot{V}O_2$ max
204 employed as the covariate - TT results are expressed as adjusted mean \pm SD or 90% CL
205 where appropriate. Standardised effect sizes (ES) were calculated to indicate the magnitude
206 of change and/or difference within- and between-groups. The criteria to interpret the
207 magnitude of ES were: <0.2 trivial, 0.2-0.6 small, 0.6-1.2 moderate, 1.2-2.0 large, and >2.0
208 very large (Hopkins, 2004).

209 Determination of biomarker concentrations and curve fit analysis was performed using
210 GraphPad Prism Version 6.03 (GraphPad Software Inc, United States) according to the
211 manufacturer's instructions. The manufacturer stated intra-assay precision was <10% for all
212 assays. Statistical analyses were performed in IBM SPSS Statistics Version 22 (IBM, United
213 States). Power analysis was conducted prior to the study and a minimum of eight participants
214 was deemed sufficient to detect the smallest worthwhile change between means assuming the
215 reference change in 5 km time trial performance was approximately twice the magnitude of
216 the typical error of measurement, with a Type I error of 5% and Type II error of 20%.

217 **Results**

218

219 *Heat Stress Test*

220 *Between group analyses*

221 . At HST₃ a significant between-group effect for TT was evident between HOT and CON
222 (HOT was faster by 8.2%, \pm 5.2%, 90% CL, $p = 0.03$). Time trial performance is presented in
223 Figure 2 as adjusted means from the analysis of covariance. No significant between-group
224 effects of short-term heat training were observed for T_c (0.3% \pm 0.6%, Figure 3), RPE,
225 TComf, sweat loss, or HR (Table 1).

226

227 *Within group analyses*

228 Both the HOT and NEUTRAL group significantly improved TT performance in HST₂ at the
229 end of the seven days short-duration protocol (after four heat training sessions) compared to
230 HST₁, with HOT 33 s \pm 20 s (adjusted mean \pm 90% CL) faster ($p = 0.02$) and NEUTRAL 39
231 s \pm 18 s faster ($p = 0.01$) than baseline. After conclusion of the post-training top-up period,
232 only HOT had a significant improvement in their TT performance at HST₃ compared to
233 HST₁, completing the course 45 s \pm 25 s faster ($p = 0.01$) compared to their HST₁
234 performance. The performance of HOT in HST₃ was also significantly improved from HST₂
235 (12 s \pm 7 s, $p = 0.01$).

236

237

238

Figure 2 about here

239 There was a small but significant mean reduction in exercising T_c observed in the HOT group
240 from HST₁ to HST₂ during the 60% workload of -0.22 ± 0.14 °C (mean \pm 90% confidence
241 limits, $p = 0.02$, ES = -0.53). Additionally, there was a trend for lower T_c during the 70%
242 workload (-0.25 ± 0.21 °C, $p = 0.06$, ES = -0.53) and during the TT (-0.25 ± 0.24 °C, $p =$
243 0.09 , ES = -0.45). Small-moderate significant reductions in T_c was observed in the HOT
244 group from HST₁ to HST₃ at the 50%; -0.18 ± 0.10 °C ($p = 0.016$), 60%; -0.23 ± 0.18 °C ($p =$
245 0.04) and 70%; -0.34 ± 0.27 °C ($p = 0.05$) workloads. The HOT group also experienced a
246 small reduction in peak T_c during HST₂ compared to HST₁; -0.25 ± 0.21 °C ($p = 0.057$), see
247 Figure 3a. Neither the NEUTRAL nor the CON group experienced meaningful reductions in
248 T_c in any of the HST's (Figure 3b and 3c).

249

250 The HOT group exhibited a moderate improvement in thermal comfort in HST₃ compared to
251 HST₁ ($p < 0.01$). Thermal comfort was also improved in HOT during HST₂ and HST₃
252 compared to NEUTRAL ($p = 0.04$ and $p = 0.03$, respectively). There were no meaningful
253 within group reductions of HR during the HST's (Table 1).

254

255 ***Figures 3 and Table 1 about here***

256

257

258

259 *Inflammatory biomarker responses*

260 *Between-group analyses*

261 No significant differences between groups in any of the biomarker responses were observed
262 either at rest or in response to any of the three HST's. Between groups there was a $\sim 8\% \pm$
263 32% difference in post HST IL-6, $\sim 52\% \pm 111\%$ in LPS, and $\sim 35\% \pm 36\%$ in IgM.

264

265 *Within-group analyses*

266 There was a large to very large ($\sim 4 \pm 2$ fold) rise in serum IL-6 concentration for all groups
267 following each HST. Serum concentrations of IgM and LPS were not substantially different
268 following the HST for each group and there were no significant time interactions observed in
269 any group. However, there was a trend for a small reduction in post-exercise concentrations
270 of IgM in all participants ($n=24$) following the first HST ($p = 0.08$, ES = 0.40). There were
271 no within-group changes observed in serum concentration of LPS ($44\% \pm 208\%$) or IgM (6%
272 $\pm 61\%$) neither pre nor post each HST. Blood biomarker concentrations are presented in
273 Figure 4.

274

275 *Figure 4 about here***

276

277

278

279 *Training sessions*

280 There were no within-group changes observed in exercising heart rate during each of the
281 training sessions for the HOT or NEUTRAL groups. Although the HOT group exhibited
282 higher HR in all training sessions compared to NEUTRAL. Table 2 outlines the physiological
283 and perceptual variables collected during the interval training sessions.

284

285 ***Table 2 about here***

286

287 **Discussion**

288 Short term heat training followed by supplementary top-up sessions (seven training sessions
289 over 18 days) improved time trial cycling performance, reduced exercising core temperature,
290 and improved thermal comfort during a strenuous cycling task in the heat. In contrast,
291 participants in the thermo-neutral (exercise) and control conditions did not experience these
292 physiological and perceptual improvements. However, as the thermo-neutral group also
293 improved their 5km TT performance after the initial short-term block of heat-training (5
294 training session in seven days), it is likely a greater stimulus in terms of intensity and
295 duration is required to elicit greater gains from heat training in shorter time periods. Although
296 mean IL-6 concentration increased four-fold following each HST, the exercise stimulus did
297 not elevate other biomarkers of systemic inflammation such as IgM and LPS. As biomarker
298 activity was largely unaffected by short-term heat training, as evidenced by IL-6 returning to
299 basal level prior to each HST, it appears that it is possible to gain useful performance and
300 thermoregulatory adaptations from short-duration training without compromising the immune
301 system. Therefore, coaches and athletes can use short-term heat acclimation training coupled
302 with supplementary heat training sessions to improve time trial performance, in the
303 confidence there is little likelihood of impairing immune system functionality.

304 Improvements in time trial performance with short-term heat training have been reported by
305 Lorenzo et al. (2010) in cycling and Garrett et al. (2012) in rowing. However Garrett and
306 colleagues did not include a control group undertaking matched training over the five day
307 heat training program. It is possible that the improvement (-4 s) observed in 2000 m rowing
308 time in that study could have been similar to that of an exercise alone control/placebo group.
309 In our study the effects of heat training were largely similar to that of the exercise-alone
310 group during the first week of training. However the supplementary top-up sessions appeared
311 to elicit further gains, indicating that while short term training offers some benefits a longer
312 program offers additional benefits. In the study by Lorenzo and colleagues, one third of the
313 experimental group (four out of twelve) were participants who had already completed the
314 control condition of the experiment, consequently, the pre-exposure to exercise in the heat
315 and heat stress test protocols. This prior exposure may have conferred a small degree of
316 acclimation prior to taking part in the experimental portion of that study. In the present study,
317 the inclusion of both an exercise matched (NEUTRAL) and control (CON) group allows
318 clear interpretation of whether the heat acclimation training was responsible for the reported
319 changes in performance and physiological adaptations. Adaptations and improvements
320 reported previously (Lorenzo et al. 2010; Garret et al. 2012) may relate to the increased
321 frequency of training within a given training period. It is likely that the heat exposure resulted
322 in ergogenic performance and thermoregulatory adaptations at the end of the 18 day period
323 beyond that of exercise training alone.

324 The improved time trial performance by participants in HOT was matched by those in
325 NEUTRAL at HST₂, indicating that the stimulus for performance gain over 7-days of short-
326 duration training in moderately-trained individuals is exercise per se rather than the
327 environmental conditions under which it is performed (i.e. hot or neutral). Although, there
328 were additional performance gains for the HOT group after the three supplementary training
329 sessions over 10 days which increased HOT's total heat load to nine exposures (two HST's
330 and seven training sessions, approx. nine hours). Clearly, exercise in temperate conditions
331 results in heat production which elevates body temperature (Gleeson, 1998), and among
332 recreationally-active participants it seems probable that this heat production is a sufficient
333 stimulus to generate modest adaptations over seven days. The observation of continued
334 adaptation and performance improvement only in the HOT group after the post-training top-
335 up period (after the full 18 days) suggests that the generic adaptive responses experienced by
336 NEUTRAL after seven days had most likely run their course and plateaued. As this study

337 recruited participants that were recreationally-active it is possible that elite endurance
338 athletes, already well-accustomed to performing regular heat producing exercise would
339 differentially experience greater gains compared to a matched neutral exercising group,
340 although this remains to be determined.

341 Although a greater number of heat exposures (than imposed in this study) could yield more
342 substantial physiological adaptations and performance improvements, it is also possible that
343 this increase could trigger systemic inflammation (Lim et al. 2009). The ~4 fold increase of
344 IL-6 concentration in all participants after the HST may not signify heat stress per se, but
345 rather the stress invoked by the exercise demand itself. IL-6 can be released into the
346 circulation following various pathological events such as physical exercise, trauma, sepsis,
347 and thermal injury (Moldoveanu et al. 2000; Natelson et al. 1996). There are few studies that
348 have investigated IL-6 as a blood biomarker during exhaustive exercise in the heat, although
349 one study reported a very large increase in IL-6 following 2 h of exhaustive walking in
350 protective clothing at 40°C (Selkirk et al. 2008). However, a different study reported a very
351 large increase in IL-6 following 3 h of exercise at 60-65% of $\dot{V}O_2$ peak in typical laboratory
352 conditions (Moldoveanu et al. 2000). Prolonged elevation of IL-6 may signify cumulative
353 fatigue or a neuroinflammatory response (Vargus et al. 2014), however in the present study
354 IL-6 returned to basal concentration prior to each HST. It appears the training load was
355 adequate to elicit some physiological and performance benefits over the 18 day period, but
356 not enough to elicit wider systemic or prolonged inflammation. Although IL-6 appeared to be
357 the most sensitive blood biomarker to the exercise task, its usefulness in specifically
358 signifying heat stress or acclimation status is limited given the non-heat specific nature of its
359 response.

360 The low concentrations of LPS observed in this study indicates the participants tolerated the
361 moderate-high heat load that was presented to them, and in doing so experienced minimal gut
362 leakage (Pyne et al. 2014). As LPS is the primary endotoxin translocated to circulation under
363 heat load (Yeh et al. 2013), its concentration and regulation is a primary consideration in
364 study of responses to the heat. It appears that undertaking ~40 min of strenuous exercise in
365 the heat is not sufficient to evoke a systemic inflammatory response in healthy moderately
366 active individuals. Furthermore, as IgM is a key antibody in neutralising LPS (Camus et al.,
367 1998), its concentration in circulating blood can reflect the body's response to endotoxin
368 accumulation and as protection against further challenges. In this study the pre- to post-
369 exercise change in IgM concentration in the heat was not significant, however following the
370 first HST there was a trend ($p=0.08$) towards reduced concentrations in all participants. It is
371 likely that a substantial heat and/or exercise stimulus may be required for IgM concentrations
372 to be substantially affected, although in this case it seems possible that there was some
373 degradation of the antibody occurring. Although some between changes were observed in
374 LPS and IgM concentrations (~44% and ~35% respectively) there was substantial uncertainty
375 in these estimates due to high variability in the biomarker response. Only one other study has
376 investigated the response of non-specific IgM following exercise in hot and humid conditions
377 (Hailes et al. 2011). During that study a 20% increase of plasma IgM was reported pre- to
378 post-exercise at day one of the heat acclimation program, this change was not present at day
379 five, with post-exercise IgM not varying from basal levels. The initial change of IgM in
380 Hailes and colleagues' study may relate to the participants required to reach a terminal core
381 temperature of 39.5 °C, whereas in the present study core temperatures did not consistently
382 rise to that level. Despite a substantial exercise and heat load (60 min HST), participants in
383 the present study were able to cope with the demands of the exercise task with limited
384 inflammation and immune disturbances.

385 **Conclusions**

386 Short-term heat training with the addition of supplementary top-up training sessions over 18
387 days enhanced time-trial performance by ~9% in recreationally-active healthy adults,
388 although thermo-neutral exercise training alone was a sufficient stimulus for performance
389 gains of ~6% over seven days. The effects of heat training appear to become more
390 worthwhile between 7-18 days. Nevertheless, training in either the heat or neutral conditions
391 proved beneficial to performance and thermoregulatory responses compared to a control
392 (non-exercise) condition. However, none of the experimental groups exhibited substantial
393 changes in LPS, IgM, or IL-6 indicating the training and heat load did not elicit an immune
394 response. It is possible that a more intense heat training protocol may lead to greater physical
395 and immune responses.

396

397 **Conflict of interest:**

398 The authors declare no conflicts of interest and this project was funded from internal funds.

399 **Acknowledgment:**

400 The authors would like to thank the participants and the laboratory staff for their time and
401 cooperation throughout this demanding study.

402

403 **Author Contributions**

404 JG, DP, GD, CM, and AE contributed to the study design. JG completed data collection and
405 conceptualization and drafting of the article. JG and KM completed Biomarker analysis. All
406 authors performed all data analysis and conceptualizing and revising the study critically for
407 important intellectual content, and approved the final manuscript

408

409 **References**

- 410 Asea, A., Kraeft, S. K., Kurt-Jones, E. A., Stevenson, M. A., Chen, L. B., Finberg, R. W., et
411 al. (2000). HSP70 stimulates cytokine production through a CD14-dependant pathway,
412 demonstrating its dual role as a chaperone and cytokine. *Nat. Med.* 6(4), 435-442. doi:
413 10.1038/74697
- 414 Borg G. (1970). Perceived exertion as an indicator of somatic stress. *Scand. J. Rehabil. Med.*
415 2, 92–98. doi:10.5271/sjweh.1815
- 416 Bouchama, A., Kwaasi, A., Dehbi, M., Al Mohanna, F., Eldali, A., El-Sayed, R., et al (2007).
417 Glucocorticoids do not protect against the lethal effects of experimental heatstroke in
418 baboons. *Shock*, 27(5), 578-583. doi: 10.1097/01.shk.0000246903.40142.aa
- 419 Camus, G., Nys, M., Poortmans, J.R., Venneman, I., Monfils, T., Deby-Dupont, G., et al.
420 1998. Endotoxaemia, production of tumour necrosis factor alpha and polymorphonuclear
421 neutrophil activation following strenuous exercise in humans. *Eur. J. Appl. Physiol.*
422 79(1), 62-68. doi: 10.1007/s004210050474
- 423 Chalmers, S., Esterman, A., Eston, R., Bowering, K. J., and Norton, K. (2014). Short-term
424 heat acclimation training improves physical performance: a systematic review, and
425 exploration of physiological adaptations and application for team sports. *Sports Med.*
426 44(7), 971-988. doi: 10.1007/s40279-014-0178-6
- 427 Daanen, H. A. M., Jonkman, A. G., Layden, J. D., Linnane, D. M., & Weller, A. S. (2011).
428 Optimising the acquisition and retention of heat acclimation. *Int. J Sports Med.* 32(11),
429 822-828. doi: 10.1055/s-0031-1279767
- 430 Dawson, B., Pyke, F. S., & Morton, A. R. (1989). Improvements in heat tolerance induced by
431 interval running training in the heat and in sweat clothing in cool conditions. *J. Sports*
432 *Sci.* 7(3), 189-203. doi: 10.1080/02640418908729840
- 433 Garrett, A. T., Goosens, N. G., Rehrer, N. G., Patterson, M. J., and Cotter, J. D. (2009).
434 Induction and decay of short-term heat acclimation. *Eur. J. Appl. Physiol.* 107(6), 659-
435 670. doi: 10.1007/s00421-009-1182-7
- 436 Garrett, A. T., Rehrer, N. J., and Patterson, M. J. (2011). Induction and decay of short-term
437 heat acclimation in moderately and highly trained athletes. *Sports Med.* 41(9), 757-771.
438 doi: 10.2165/11587320-000000000-00000
- 439 Garrett, A. T., Creasy, R., Rehrer, N. J., Patterson, M. J., and Cotter, J. D. (2012).
440 Effectiveness of short-term heat acclimation for highly trained athletes. *Eur. J. Appl.*
441 *Physiol.* 112(5), 1827-1837. doi: 10.1007/s00421-011-2153-3
- 442 Gibson, O. R., Mee, J. A., Taylor, L., Tuttle, J. A., Watt, P. W., and Maxwell, N. S. (2015).
443 Isothermic and fixed-intensity heat acclimation methods elicit equal increases in Hsp72
444 mRNA. *Scand. J. Med. Sci. Sport.* 25(S1), 259-268. doi: 10.1111/sms.12430
- 445 Gill, S. K., Teixeira, A., Rama, L., Prestes, J., Rosado, F., Hankey, J., and Costa, R. J. (2014).
446 Circulatory endotoxin concentration and cytokine profile in response to exertional-heat
447 stress during a multi-stage ultra-marathon competition. *Exerc. Immunol Rev.* 21, 114-128.
- 448 Gleeson, M. (1998). Temperature regulation during exercise. *Int. J. Sports Med.* 19, S96-9.
449 doi: 10.1055/s-2007-971967

- 450 Guy, J. H., Deakin, G. B., Edwards, A. M., Miller, C. M., and Pyne, D. B. (2015). Adaptation
451 to hot environmental conditions: an exploration of the performance basis, procedures and
452 future directions to optimise opportunities for elite athletes. *Sports Med.* 45(3), 303-311.
453 doi: 10.1007/s40279-014-0277-4
- 454 Hailes, W. S., Slivka, D., Cuddy, J., and Ruby, B. C. (2011). Human plasma inflammatory
455 response during 5 days of exercise training in the heat. *J. Therm. Biol.* 36(5), 277-282. doi:
456 10.1016/j.jtherbio.2011.03.013
- 457 Hopkins, W.G. 2004. How to interpret changes in an athletic performance test. *Sportscience*,
458 8, 1-7. sportsci.org/jour/04/wgtests.htm.
- 459 Harding S. (2011). The tropical agenda. *J. Trop. Psychol.* 1(1), 2-5. doi: 10.1375/jtp.1.1.2
- 460 Kelly, M., Gastin, P. B., Dwyer, D. B., Sostaric, S., & Snow, R. J. (2016). Short duration heat
461 acclimation in Australian football players. *J. Sports Sci. Med.* 15(1), 118.
- 462 Kregel, K. C. (2002). Invited review: heat shock proteins: modifying factors in physiological
463 stress responses and acquired thermotolerance. *J. Appl. Physiol.* 92(5), 2177-2186. doi:
464 10.1152/jappphysiol.01267.2001
- 465 Lau, S. S., Griffin, T. M., and Mestrlil, R. (2000). Protection against endotoxemia by HSP70
466 in rodent cardiomyocytes. *Am. J. Physiol-Heart C.* 278(5), H1439-H1445.
- 467 Lorenzo, S., Halliwill, J. R., Sawka, M. N., and Minson, C. T. (2010). Heat acclimation
468 improves exercise performance. *J. Appl. Physiol* 109(4), 1140-1147. doi:
469 10.1152/jappphysiol.00495.2010
- 470 Nielsen, B., Strange, S., Christensen, N. J., Warberg, J., & Saltin, B. (1997). Acute and
471 adaptive responses in humans to exercise in a warm, humid environment. *Pflügers*
472 *Archiv*, 434(1), 49-56. doi: 10.1007/s004240050361
- 473 Magalhães, F. C., Amorim, F. T., Passos, R. L. F., Fonseca, M. A., Oliveira, K. P. M., Lima,
474 M. R. M., and Rodrigues, L. O. C. (2010). Heat and exercise acclimation increases
475 intracellular levels of Hsp72 and inhibits exercise-induced increase in intracellular and
476 plasma Hsp72 in humans. *Cell Stress Chaperon.* 15(6), 885-895. doi: 10.1007/s12192-
477 010-0197-7
- 478 Magalhães, F. C., Passos, R. L., Fonseca, M. A., Oliveira, K. P., Ferreira-Júnior, J. B.,
479 Martini, A. R., and Rodrigues, L. O. (2010). Thermoregulatory efficiency is increased
480 after heat acclimation in tropical natives. *J. Physiol. Anthropol.* 29(1), 1-12. doi:
481 10.1007/s12192-010-0197-7
- 482 Midgley, A. W., McNaughton, L. R., Polman, R., and Marchant, D. (2007). Criteria for
483 determination of maximal oxygen uptake. *Sports Med.*, 37(12), 1019-1028. doi:
484 10.2165/00007256-200737120-00002
- 485 Moldoveanu, A.I., Shephard, R.J., and Shek, P.N. (2000). Exercise elevates plasma levels but
486 not gene expression of IL-1 β , IL-6, and TNF- α in blood mononuclear cells. *J. Appl.*
487 *Physiol.* 89(4), 1499-1504. doi:
- 488 Peake, J. (2010). Heat, Athletes, and Immunity. *Am. J. Lifestyle Med.* 4(4), 320-326. doi:
489 10.1177/1559827610362969
- 490 Périard, J. D., Racinais, S., and Sawka, M. N. (2015). Adaptations and mechanisms of human
491 heat acclimation: applications for competitive athletes and sports. *Scand. J. Med. Sci.*
492 *Sport.* 25(S1), 20-38. doi: 10.1111/sms.12408

- 493 Pyne, D.B., Guy, J.H., and Edwards, A.M. 2014. Managing Heat and Immune Stress in
494 Athletes with Evidence-Based Strategies. *Int. J. Sports Physiol. Perform.* 9(5), 744-750.
495 doi: 10.1123/ijsp.2014-0232
- 496 Sawka, M. N., Wenger, C. B., and Pandolf, K. B. (2011). Thermoregulatory responses to
497 acute exercise-heat stress and heat acclimation. *Compr. Physiol.* doi:
498 10.1002/cphy.cp040109
- 499 Selkirk, G.A., McLellan, T.M., Wright, H.E., and Rhind, S.G. 2008. Mild endotoxemia, NF-
500 κ B translocation, and cytokine increase during exertional heat stress in trained and
501 untrained individuals. *Am. J. Physiol. Regul. Integr. Compr. Physiol.* 295(2), R611-R623.
502 doi: 10.1152/ajpregu.00917.2007
- 503 Vargas, N. T., & Marino, F. (2014). A neuroinflammatory model for acute fatigue during
504 exercise. *Sports Med.* 44(11), 1479-1487. doi: 10.1007/s40279-014-0232-4
- 505 Wingfield, G. L., Gale, R., Minett, G. M., Marino, F. E., & Skein, M. (2016). The effect of
506 high versus low intensity heat acclimation on performance and neuromuscular responses.
507 *J. Therm. Biol.* 58, 50-59. doi: 10.1016/j.jtherbio.2016.02.006
- 508 Yeh, Y.J, Law, L.Y., and Lim, C.L. 2013. Gastrointestinal response and endotoxemia during
509 intense exercise in hot and cool environments. *Eur. J. Appl. Physiol.* 113(6), 575-1583.
510 doi: 10.1007/s00421-013-2587-x

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Figure 1. Study timeline for Heat Training (HOT), Thermo-neutral Training (NEUTRAL) and Control (CON) groups.

Figure 2. Adjusted means \pm SD of 5km time trial performance (s) across heat stress tests (HST) 1, 2 and 3 for Heat (HOT), Thermo-neutral (NEUTRAL) and Control (CON) groups. * Faster from baseline. † Faster than HST 2. Ω HOT was faster than CON.

Figure 3. Core temperature for Heat Training (HOT), Thermo-neutral Training (NEUTRAL) and Control (CON) groups during Heat Stress Tests (HST) 1, 2, and 3, expressed as mean \pm SD. * Reduced from baseline at HST 2. † Reduced from baseline at HST 3.

Figure 4. Serum concentrations of interleukin 6 (IL-6), Immunoglobulin M (IgM) and Lipopolysaccharide pre and post Heat Stress Tests 1, 2, and 3. * Increased from pre exercise concentration.

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Table 1. Physiological and perceptual responses to Heat Stress Tests

	HST ₁			HST ₂			HST ₃		
	HOT	NEUTRAL	CON	HOT	NEUTRAL	CON	HOT	NEUTRAL	CON
HR _{50%} (bpm)	139 ± 15	135 ± 12	137 ± 14	136 ± 15	133 ± 11	138 ± 13	136 ± 17	133 ± 10	133 ± 13
HR _{60%} (bpm)	162 ± 15	159 ± 9	157 ± 9	155 ± 14	154 ± 9	156 ± 9	155 ± 16	154 ± 11	153 ± 11
HR _{70%} (bpm)	175 ± 13	178 ± 7	170 ± 8	169 ± 13	172 ± 9	170 ± 6	168 ± 13	171 ± 9	167 ± 7
HR _{TT} (bpm)	177 ± 11	178 ± 9	169 ± 10	176 ± 9	179 ± 6	168 ± 7	179 ± 10	175 ± 10	164 ± 12
RPE _{Avg} (units)	14 ± 1	14 ± 1	15 ± 1	13 ± 2	14 ± 2	13 ± 1	13 ± 2	15 ± 3	13 ± 2
RPE _{End} (units)	17 ± 2	17 ± 2	17 ± 2	17 ± 2	18 ± 2	17 ± 3	17 ± 2	17 ± 2	16 ± 3
TComf _{Avg} (units)	3.0 ± 0.5	3.0 ± 0.5	3.5 ± 0.5	2.0 ± 1.0*	3.0 ± 0.5	3.0 ± 1 ^Ω	2.0 ± 1.0* [†]	3.0 ± 0.5 [∞]	3.0 ± 0.5* ^Ω
TComf _{End} (units)	4.0 ± 0.5	4.5 ± 0.5	4.5 ± 0.5	3.0 ± 1.0	4.5 ± 1.0 [∞]	4.0 ± 1	3.0 ± 1.0*	4.0 ± 1.0	3.5 ± 1.0

Data are expressed as mean ± SD. HOT = Heat training group, NEUTRAL = Thermo-neutral training group, CON = Control group. HR = Heart rate. Sweat loss (%) is expressed as the amount of sweat lost (kg) divided by the persons pre-exercise mass (kg) x 100. RPE_{Avg} and TComf_{Avg} are the mean Rating of Perceived Exertion and Thermal Comfort rating across the entire Heat Stress Test (HST). RPE_{End} and TComf_{End} represent the values recorded at the cessation of the HST. *Significantly different from HST₁. [†] Significantly different from HST₂. [∞] Significant difference between HOT and NEUTRAL. ^Ω Significant difference between HOT and CON.

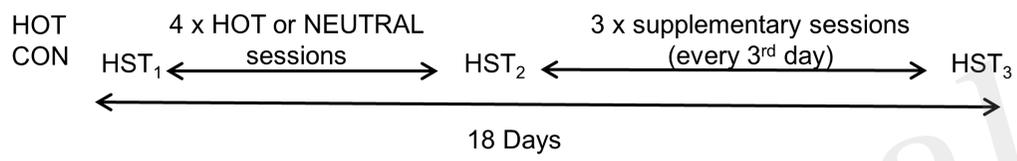
Table 2. Physiological and perceptual observations during sub-maximal aerobic interval training from training sessions one, four, and the third top up session

	TR ₁		TR ₄		TU ₃	
	HOT	NEUTRAL	HOT	NEUTRAL	HOT	NEUTRAL
HR (bpm)	161 ± 13	145 ± 9 [∞]	157 ± 12	145 ± 6 [∞]	154 ± 15	140 ± 13
RPE _{Avg} (units)	15 ± 1	15 ± 2	14 ± 2	15 ± 2	13 ± 3	13 ± 1 [†]
TComf _{Avg} (units)	3.0 ± 1.0	3.0 ± 1.0	3.0 ± 1.0	3.0 ± 1.0	2.0 ± 1.0	3.0 ± 1.0

Data is expressed as mean ± SD. HOT = Heat training group, NEUTRAL = Thermo-neutral training group. TR₁ = Training session on day one, TU₃ = Top up training session on day 15. HR = Mean heart rate across four x 10 minute intervals. RPE_{Avg} and TComf_{Avg} are the mean Rating of Perceived Exertion and Thermal Comfort rating across the training session. * Significantly different from TR₁. † Significantly different from TR₄. ∞ Significant difference between HOT and NEUTRAL.

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Figure 01.TIF



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Figure 02.TIF

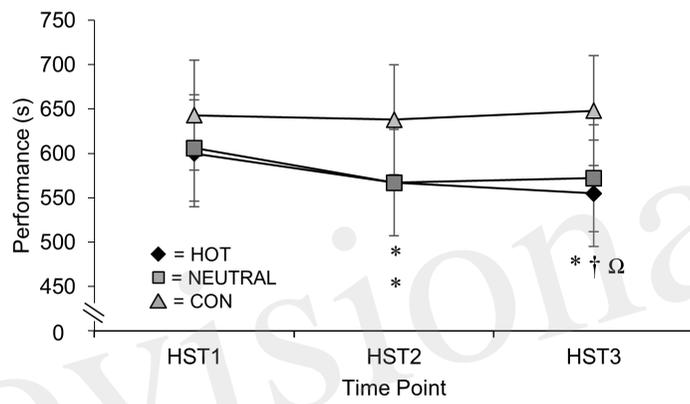


Figure 03.TIF

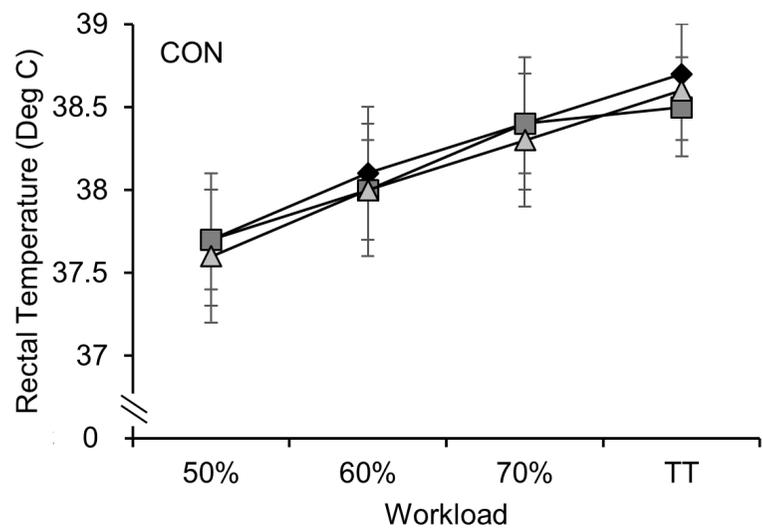
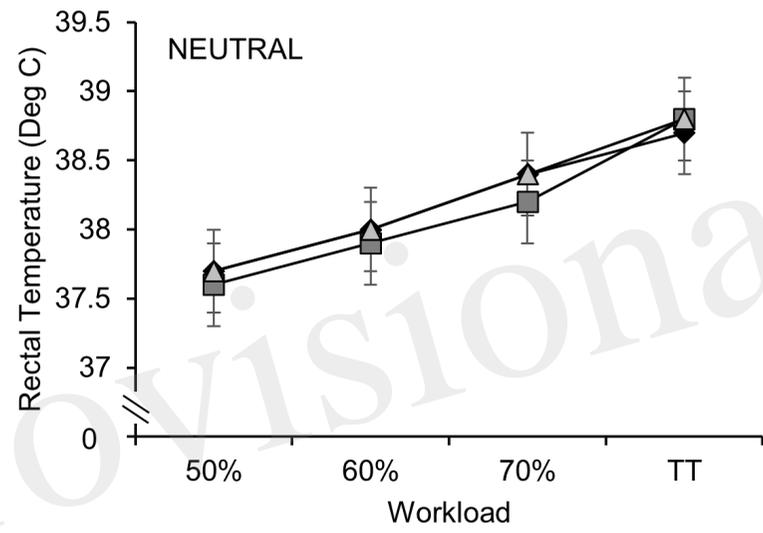
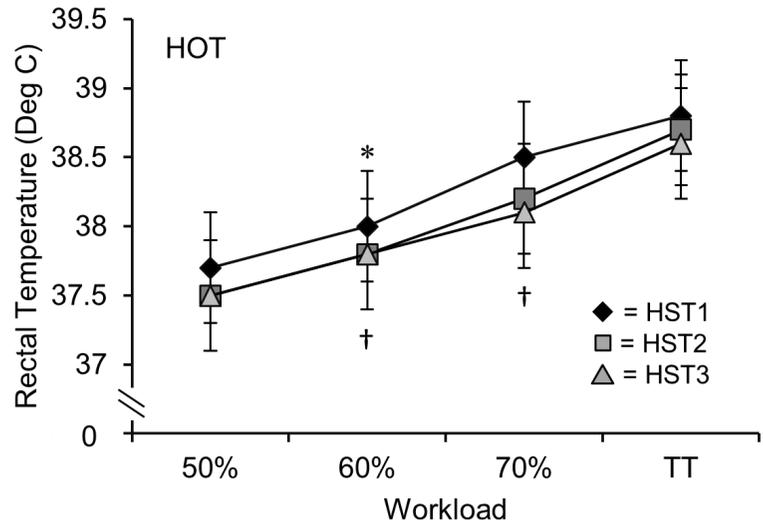


Figure 04.TIF

